Xylanase and xylo- oligosaccharide prebiotic improve the growth performance and concentration of potentially prebiotic oligosaccharides in the ileum of broiler chickens

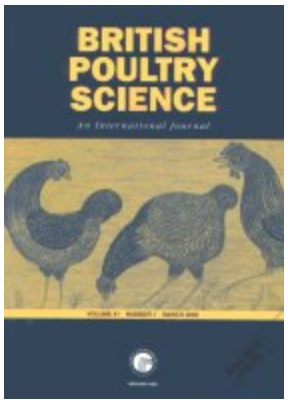
Craig, AC; Khattak, FM; Hastie, P; Bedford, Michael R; Olukosi, OA

Published in:
British Poultry Science

DOI:
10.1080/00071668.2019.1673318

Print publication: 01/02/2020

Document Version
Peer reviewed version



Xylanase and xylo- oligosaccharide prebiotic improve the growth performance and concentration of potentially prebiotic oligosaccharides in the ileum of broiler chickens

Journal:	*British Poultry Science*
Manuscript ID	CBPS-2019-183.R1
Manuscript Type:	Original Manuscript
Date Submitted by the Author:	05-Aug-2019
Complete List of Authors:	Craig , Allison ; Scotland's Rural College, Monogastric Science Research Centre; University of Glasgow, McCall Building, School of Veterinary Medicine Khattak, Farina ; Scotland's Rural College, Monongastric Science Research Centre Hastie, Peter ; University of Glasgow, McCall Building, School of Veterinary Medicine Bedford, Michael; ABVista, Woodstock Centre, Marlborough Business Park Olukosi, Oluyinka; Scotland's Rural College; University of Georgia, Department of Poultry Science
Keywords:	Broilers, Xylanase, Nutrient digestibility, Xylose oligosaccharides, short chain fatty acids, Growth

Xylanase and xylo-oligosaccharide prebiotic improve the growth performance and concentration of potentially prebiotic oligosaccharides in the ileum of broiler chickens.

A. D. Craig[1, 2]., F. Khattak[1]., P. Hastie[2]., M.R. Bedford[3]., and O. A. Olukosi[1,4*].

[1] Monogastric Science Research Centre, SRUC, Edinburgh, EH9 3JG

[2]McCall Building, School of Veterinary Medicine, University of Glasgow, G61 1QH

[3]A B Vista, Woodstock Centre, Marlborough Business Park, Marlborough, SN8 4AN

[4] Department of Poultry Science, University of Georgia, Athens, GA 30602, USA

[*]Corresponding author

Oluyinka A. Olukosi, Department of Poultry Science, University of Georgia, Athens, GA 30602 Email: oaolukosi@uga.edu Tel: +1 706 542 1359

ORCID- Miss A.D Craig – 0000-0002-1640-8612

ORCID- Dr O.A Olukosi- 0000-0002-9137-6745

Accepted for publication 22 August 2019

ABSTRACT

1. The objective of this study was to investigate the effect of supplementing broiler diets with xylanase or xylo- oligosaccharide (XOS) on growth performance, the concentration of non-starch polysaccharide (NSP) hydrolysis products in the ileum and concentration of short chain fatty acids (SCFA) in the caeca of broiler chickens.
2. In total, 500 male Ross 308 broilers were used in this 29-day (d) study. The treatments were organised into a 2×2 plus 1 factorial arrangement consisting of two additives (xylanase or XOS) at two levels (low or high) plus a control treatment with no additives. This gave five treatments with 100 bird in each treatment group. The diets were slightly deficient in protein by 20 g/kg and energy by 1 MJ/kg.
3. On d 14 and 28, two birds per pen were euthanised, the caeca content collected and analysed for short chain fatty acid (SCFA) concentration. On d 29, six birds per pen were euthanised and ileal digesta were collected and analysed for the concentration of NSP fractions.
4. On d 14, caecal acetic acid, iso-butyric acid, iso-valeric acid, n-valeric acid and total SCFA concentrations were significantly greater ($P \leq 0.05$) when diets were supplemented with XOS compared with xylanase.
5. Ileal concentration of arabinose, galactose and glucuronic acid (GlucA2) were significantly greater ($P \leq 0.05$) in the insoluble NSP fraction when diets were supplemented with a high level of xylanase, compared with the control treatment. Ileal concentration of fructose was significantly greater ($P \leq 0.05$) in the water soluble NSP when a high level of xylanase or low level of XOS were included in the diet compared with the control.

Accepted for publication 22 August 2019

6. It was concluded that xylanase and XOS had similar effects on NSP concentration and SCFA in the caeca, although there was little effect on performance. This observation demonstrated further benefits of xylanase supplementation in wheat-based broiler diets beyond digesta viscosity reduction and the release of extra nutrients.

Keywords- xylanase, XOS, broilers, performance, nutrient digestibility, SCFA

Accepted for publication 22 August 2019

Introduction

Depression in growth performance caused by an increase in digesta viscosity is a common occurrence in broiler diets containing a large amount of non-starch polysaccharides (NSP; Jia *et al.*, 2009). In order to overcome this, carbohydrase enzymes are often added to broiler diets to improve nutrient utilisation and increase productivity. Carbohydrases hydrolyse NSP, breaking it down into smaller oligosaccharides. This results in a decrease in digesta viscosity and the release of encapsulated nutrients (Knudsen, 2014). In addition to these benefits, it has been suggested that the small oligosaccharides produced during NSP hydrolysis could have prebiotic properties (Courtin *et al.*, 2008).

One way of investigating the production of small oligosaccharides is to measure the concentration of NSP hydrolysis products in the ileum of broilers, such as arabinose or xylose concentrations. A prebiotic is a small molecule which is fermented by beneficial bacteria, encouraging their growth, while discouraging the growth of pathogenic bacteria. Xylo-oligosaccharides (XOS) are associated with improvements in poultry performance (Al-Sultan *et al*., 2016) by modulating the gastrointestinal immune system, microbial populations (Jung *et al.*, 2008; Yang *et al.,*2008) and increasing short chain fatty acid (SCFA) production, however, evidence for this has been inconsistent (Arsi *et al.,*2015; Gao *et al.*, 2007).

To investigate the production of potentially prebiotic oligosaccharides during xylanase activity, monosaccharides were measured in the ileal digesta and compared to monosaccharides in digesta from birds supplemented with purified XOS. The objective of the trial was to investigate similarities in profiles of monosaccharides in

Accepted for publication 22 August 2019

digesta from birds receiving xylanase and those receiving XOS to illustrate that NSP hydrolysis products may have prebiotic-like effects similar to that of purified XOS.

NSP hydrolysis products are thought to be fermented by beneficial bacteria such as *Bifidobacter* and *Lactobacilli spp.,* producing SCFA (Lee *et al.,* 2017). An increase in the concentration of SCFAs is often associated with an increase in the population of beneficial bacteria and a decrease in pathogenic bacteria (Engberg *et al.,* 2004). In addition to this, SCFAs have been shown to influence growth performance in broilers. Butyrate, in particular, is regarded as an available energy source, increasing the energy available to the host for growth (Ravangard *et al.,* 2017). Supplementing broiler diets with xylanase or XOS has been shown to affect the production of SCFA in the caeca of broiler chickens (Engberg *et al.,* 2004; Lee *et al.,* 2017). This could suggest that both ingredients have a similar mode of action. As such, the effect of xylanase or XOS on SCFA concentration in the caeca of broilers was investigated in the current study.

The objective of this experiment was to investigate the effect of supplementing wheat-based diets, which were deficient in energy and protein, with xylanase or XOS on growth performance, the concentration of NSP hydrolysis products in the ileum and the concentration of SCFA's in the caeca of broiler chickens.

Materials and methods

Animals and management

All the procedures in the experiment were approved by the SRUC Animal Experiment Committee.

Accepted for publication 22 August 2019

Five hundred male Ross 308, one-d-old broilers were allocated to one of five treatments organised as a randomised complete block design. The birds were housed 10 in a pen, with ten pen replicates per treatment and provided with feed and water on an *ab libitum* basis throughout the experiment (0 to 29 d). The treatments followed a 2×2 plus 1 factorial arrangement with two different additives (xylanase or XOS) at two inclusion levels (high and low) plus the control. The low level of xylanase inclusion was 16, 000 XU/kg and the high was 32,000 XU/kg. The low level of XOS inclusion was 0.25 g/kg and the high level was 0.1 g/kg. The lower level of each additive was based on the standard recommendation by the manufacturer while the higher level of each additive was chosen following a literature search (Wang *et al*., 2005; Zhenping *et al.,* 2013; De Measschalck *et al*., 2015). The control diet was deficient in energy by 1 MJ/kg and protein reduced by 3%, to 20g/kg CP however; all other nutrient requirements were met to ensure that any effects were induced by energy or protein deficiency or additive supplementation alone. The diets were formulated to be deficient in energy and protein to allow any improvements in growth or nutrient digestibility to become apparent, as previous work has indicated that xylanase supplementation may be more beneficial in nutrient-deficient diets (Francesch and Geraert, 2009). The experiment was split into five treatments; 1) control, 2) control plus xylanase 16,000 XU/kg, 3) control plus xylanase 32,000 XU/kg, 4) control plus purified XOS 0.25 g/kg and 5) control plus purified XOS 1.0 g/kg. All of the experimental diets were provided in mash form and birds had *ad libitum* access to feed and water throughout the feeding trial. The xylanase used contained 160 000 U of endo- 1,4 β xylanase activity per gram. One unit of xylanase activity was defined as the amount of enzyme required to liberate 1 µmol of reducing sugars from xylan using a standardised test (Enzyme

Accepted for publication 22 August 2019

Services (ESC), Hengoed, Ystrad Mynack, UK). The xylanase (Econase XT) was supplied by AB Vista, Marlborough, UK. The XOS used in this study was purchased from Shandong Lifelong Bio-technology Co., China (XOS 35A). The ingredient and chemical composition of the control diet is shown in Table 1. Wheat bran was included in the diet during this study to increase the amount of NSP in the diet and maximise the potential for prebiotic oligosaccharide generation during NSP hydrolysis by xylanase.

Table 1 here

Growth performance

Feed and birds were weighed on d 0, 14 and 28. The data from feed and bird weights were used to calculate body weight gain (BWG), feed intake (FI) and feed conversion ratio (FCR).

Sample collection

On d 14 and 28, two birds per pen were euthanised by cervical dislocation and used to collect caeca content for SCFA analysis. Following euthanasia, the caeca were removed and the content was gently squeezed into a collection tube.

On d 29, the remaining six birds per pen were euthanised by cervical dislocation and ileal content was collected for nutrient digestibility and NSP analysis. Once located, the distal half of the ileum was removed and the contents were flushed with water into a collection pot. The ileal digesta from all six birds per pen was pooled and was

collected on day 29, as opposed to day 28, due to practical reasons relating to volume of samples to be collected.

Short chain fatty acid analyses

Once the contents of the caeca were removed they were stored at -20^{0}C and later analysed for SCFA concentration as described by Khattak *et al*. (2018) using gas chromatography.

NSP fraction analyses

The ileal digesta samples collected from six birds per pen on d 29 were dried in a Unitherm force draft drying oven for 48 hours at 80^{0}C. The samples were analysed for water extractable (WE) carbohydrate components and water unextractable (WU) NSP using HPLC, following the method of Englyst *et al.* (1994). The pre-caecal NSP concentration was calculated using the equation displayed in the calculations section.

Chemical analysis

The ileal digesta samples collected on d 29 were dried prior to conducting titanium and DM analysis according to the method of Short *et al.* (1996). Dry matter (DM) was determined using standard methods from AOAC, whereby 1g of the sample was dried in a uniform forced drying oven (Unitherm, Russel-Lindsay Engineering Ltd, Birmingham, England, UK) at 95°C for 24 hours. Nitrogen determination was carried out by the combustion method (Method 934.01; AOAC). Gross energy was determined using an isoperibol bomb calorimeter system using benzoic acid as an internal standard (Model 6200, Parr Instruments, Moline, Illinois, USA). Ileal digestibility was calculated using the index method described by Olukosi *et al.*

Accepted for publication 22 August 2019

(2007). The activity of xylanase in the diet was measured using a commercial test kit (Enzyme Services (ESC), Hengoed, Ystrad Mynack, UK). One BXU was defined as the amount of xylanase enzyme required to liberate 1 nmol of reducing sugars per minute from xylan at pH 5.3.

Calculations

Pre-caecal NSP concentration (g/100g DM intake) was calculated using the equation below:

$$NSP\ Concentration = NSP\ conc.\ in\ digesta \times \left(\frac{Ti\ in\ diet}{Ti\ in\ digesta}\right)$$

NSP Conc. In digesta (%)

Statistics

Statistical analysis was conducted using the ANOVA function of Genstat (16th Edition). Data were analysed following the 2 2 plus 1 factorial arrangement with body weight as the blocking factor. When an interaction between additive type and inclusion level (excluding the no additive treatment) was significant, the means for growth performance and nutrient digestibility were separated using Tukey's test. The significant additive type × inclusion level interactions for ileal NSP concentration were detected using specific contrasts details of which are in Tables 5 and 6. Significance was set at $P \leq 0.05$ and trends at $P < 0.1$.

Results

Growth performance of broilers on days 14 and 28

Accepted for publication 22 August 2019

The enzyme analysis results showed that diets 1, 4 and 5 contained xylanase activity below detection threshold. Diet 2 contained 16,600 BXU/kg and diet 3 contained 36,900 BXU/ kg of xylanase activity.

On d 14 there was no additive type×inclusion level interaction for BWG, FI or FCR both on d 14 and 28 (Table 2). However, there were main effects of additive type and inclusion level on d 14. Body weight gain and feed intake were greater ($P<0.05$) following xylanase supplementation compared with XOS supplementation. Feed conversion ratio was lower ($P<0.05$) following additive supplementation at high inclusion level compared with low inclusion level. There was no effect of additive supplementation compared with the unsupplemented control.

On d 28, there were no significant ($P<0.05$) main or interaction effects of additive type or inclusion level for BWG, FI or FCR. However, feed intake and FCR values were lower ($P<0.001$) for broilers receiving xylanase or XOS compared with the control.

Table 2 here

Nutrient digestibility in broilers aged 29 days and fed diets supplemented with xylanase or XOS

Nutrient digestibility in broilers aged 29 d and fed diets supplemented with xylanase or XOS is shown in Table 3. There was a significant additive type×inclusion level interaction for nitrogen (N) intake. Birds receiving diets containing the low level of xylanase had significantly ($P<0.01$) lower N intake than those receiving the high

Accepted for publication 22 August 2019

level of xylanase. There was no effect of additive type or inclusion level on DM or N digestibility, however, there was trend for greater ileal digestible energy (IDE) in birds fed diets supplemented with xylanase compared to those with XOS. The DM, IDE and N digestibilities were lower (P<0.05) in birds fed diets containing xylanase or XOS compared to the control. The N and gross energy intake were lower (P<0.05) in birds fed diets containing xylanase or XOS compared to the control diets.

Table 3 here

Short chain fatty acid concentration in the caeca of broiler chickens on days 14 and 28

Caecal SCFA concentration in response to xylanase or XOS supplementation on d 14 is shown is Table 4. There was no significant inclusion level or additive type×inclusion level interaction for SCFA concentration on d 14, however there was a main effect of ingredient type. The concentration of acetic acid, propanioc acid, iso-butyric acid, iso- valeric acid and total SCFA were greater (P<0.05) following XOS compared with xylanase supplementation. There was no effect of ingredient inclusion compared to the control treatment on acetic acid, n-butyric acid or n-valeric acid concentration in broilers on d 14. However, there was a trend (P=0.066) for xylanase to decrease propionic acid concentration and XOS to increase propionic acid concentration.

Caecal SFCFA concentration in response to xylanase or XOS supplementation on d 28 is shown in Table 4. There was no additive type×inclusion level interaction for

SCFA concentration on d 28. There was no main effect of ingredient type or inclusion level on caecal SCFA concentration on day 28.The concentration of acetic acid, n-butyric acid and total SCFA were greater ($P<0.05$) following ingredient supplementation compared to the control. The concentration of acetic acid, propanioc acid and total SCFA was greater ($P<0.05$) in birds aged 28 days compared to birds aged 14 days.

Table 4 here

NSP fraction content of ileal digesta from broiler chickens aged 29 days

The concentration of WU NSP fractions in response to xylanase or XOS supplementation is shown in Table 5. There was an ingredient type×inclusion level interaction for arabinose and galactose. WU arabinose and galactose concentration were greater when diets were supplemented with the high level of xylanase compared to the low level. When the control treatment was compared to the other treatments individually, arabinose and galactose concentration were greater ($P<0.05$) following xylanase supplementation at a high level, compared to the control. There were main effects of ingredient type and inclusion levels. Rhamnose concentration was greater ($P<0.001$) following XOS compared to xylanase supplementation. Rhamnose and fructose concentration were greater ($P<0.05$) following supplementation at a high level compared to the low level. The concentration of rhamnose, fructose, arabinose and galactose were greater ($P<0.05$) following supplementation compared with the control.

Accepted for publication 22 August 2019

Table 5 here

The concentration of WE NSP fractions in response to xylanase or XOS supplementation is shown in Table 6. There was a significant ingredient type×inclusion level interaction for fructose concentration. The WE fructose concentration was greater when the high level of xylanase or the low level of XOS were included in the diet, compared to the low level of xylanase or high level of XOS. When the control treatment was compared to the other diets individually, WE fructose concentration was greater ($P<0.05$) following supplementation of 32,000 XU/kg xylanase or 0.025% XOS compared to the control. There were no main effects of ingredient type or inclusion level for WE NSP concentration. There was a tendency ($P=0.061$) for greater xylose concentration in diets supplemented with XOS compared to xylanase. There was a tendency for greater galactose concentration in diets supplemented with the high compared to the low inclusion level. Galactose and total WE NSP concentration were greater ($P<0.05$) following supplementation compared with the control.

Table 6 here

Discussion

Accepted for publication 22 August 2019

Xylanase is used routinely in broiler diets to improve growth performance, however, there is evidence to suggest that potentially prebiotic oligosaccharides are generated during NSP hydrolysis. The generation of *in-situ* prebiotics could provide additional benefit to the use of xylanases in broiler diets over and above the reduction in digesta viscosity.

Growth Performance

The birds in the current study performed below breed standards. This could be due to a multitude of reasons, including diet form and composition. It has been well established in the literature that low protein (23 -20% CP) or low energy (3000-2640 Kcal/kg) diets reduce the growth performance of broilers (Govil *et al.,* 2017; Kamran *et al.,* 2008; Williams *et al.,* 2014). The current study is in agreement with this, as the birds receiving the control feed, which was lower in energy by 8% and protein by 13%, ate significantly more than those birds receiving supplementation in their diet. This was expected to an extent, as it has been suggested that birds do not eat to satisfy hunger *per se* but they consume enough feed to satisfy their energy or protein requirements (Karam *et al.,*2008). In the current study, the birds receiving diets low in energy and protein were able to increase their feed intake, allowing them to maintain their growth. When xylanase or XOS was added into the diet, feed intake reduced, as expected, resulting in a reduction in FCR and an increase in efficiency.

Xylanase supplementation improved BWG, FI and FCR compared to that of XOS on d 14. This was expected as the mechanism of action of xylanase is well established in the literature. Xylanase cleaves the arabinoxylan (AX) backbone of NSP releasing the trapped nutrients (Meng *et al*., 2005) and reduces digesta viscosity (Lentle, 2005). This impacts on growth performance in two ways. Firstly, the release of

Accepted for publication 22 August 2019

trapped nutrients increases the amount available for absorption in the small intestine (Meng *et al.,* 2005). Secondly, reducing digesta viscosity allows sufficient mixing of the digesta, enabling more nutrients to be absorbed (Lentle, 2005). There is evidence to suggest a third mechanism, namely generation of *in-situ* prebiotic oligosaccharides. During the hydrolysis of NSP, smaller oligosaccharides such as XOS are generated which have been shown to have prebiotic-like effects (Zhang *et al.,* 2014).

Prebiotics can improve growth performance of broilers (Al-Sultan *et al.,* 2016; Abdel- Hafeez *et al.,* 2017). In the current study, on d 28, XOS reduced feed intake and improved FCR but had no effect on body weight gain, which was similar to the effect of xylanase supplementation. It is thought that prebiotics improve growth performance by increasing nutrient absorption due to modulating gut microflora and increasing gut integrity (Al-Sultan *et al.,* 2016). Beneficial gut bacteria, such as *Bifidobacterium* and *Lactobacilli spp.,* ferment prebiotics, such as XOS, FOS and GOS, which encourages growth whilst discouraging the colonisation of pathogenic bacteria (Xu *et al.*, 2003; Courtin *et al.,* 2008; Yousaf *et al.,* 2016). This may relate to the SCFA results, discussed below.

Nutrient digestibility

Ingredients such as xylanase have been shown to improve nutrient digestibility (Kiarie *et al.,* 2014). The expectation that adding carbohydrases can improve nutrient digestibility is logical, as the enzyme should decrease digesta viscosity and allow increased absorption of nutrients (Mathlouthi *et al.,* 2002). In the current study, DM and N digestibility were reduced when either xylanase or XOS were added to the diet, however, IDE was significantly lower when XOS was included in the diet. The

diets used during this study were deficient in energy and protein, so the aim of adding such ingredients was to improve nutrient absorption, as this is what has been described in the literature (Cowieson *et al.,* 2017). The addition of xylanase or XOS reduced digestible energy and N intake compared to the control treatment, which may help explain why nutrient digestibility was decreased following supplementation.

The negative effect of XOS supplementation on IDE was unexpected. The literature often reports no effect of prebiotics on ileal nutrient digestibility (Kirkpinar *et al.,*2004; Mountzouris *et al.,* 2010) however improvements in total tract retention have been reported (Mountzouris *et al.,* 2010). This is achieved when populations of beneficial microflora are encouraged, which increases nutrient digestion and adsorption (De Measschalck *et al.,* 2015) of SCFAs produced during fibre fermentation. Decreased nutrient digestibility following prebiotic supplementation has been reported previously in pigs. The authors suggested that the reduction in nutrient digestibility was due to the introduction of indigestible fibre into the diet (Smiricky- Tjardes *et al.,* 2003). This is an unlikely explanation for the reduction in nutrient digestibly reported in the current study, however, the inclusion level of XOS was much lower that than that used by Smiricky- Tjardes *et al.,* (2003).

There was an effect of supplementation on nutrient digestibility however there was little effect on growth performance especially on d 28, meaning that the improvements in nutrient digestibility were not translated into growth performance, which has been reported previously (Yang *et al*., 2008; Gonzalez- Ortiz *et al*., 2016). This could indicate that 'point in time' measurements, such as nutrient digestibility, are poor tools in predicting the long-term effects of a diet on performance parameters such as body weight gain or FCR.

Accepted for publication 22 August 2019

Short chain fatty acid concentration

Bird age had a significant effect on the concentration of SCFAs in the caecum of broiler chickens. On d 28, there was a greater concentration of acetic, propionic, iso-butyric and iso-valeric acid compared to SCFA on d 14. This is in agreement with Lee *et al.* (2017). The authors showed that the concentration of SCFAs increased as the bird aged. The reason for this was likely to be the development of the intestinal microflora. Gong *et al.,* (2008) demonstrated that young birds (14 d of age) had a less well-developed microflora than those aged 42 d. Not only was the microflora in 14 d old birds less well developed, it was more likely to be influenced by changes in the diet or environment. In the current study, it was possible that SCFA concentration in the caeca were lower on d 14, because the microflora were less well developed and not able to ferment carbohydrate sources in order to produce SCFA. As the bird aged, the microflora matured and established itself, enabling the microbes to more readily ferment available carbohydrates, increasing the production of SCFA, which is in agreement with Lee *et al.,* (2017).

The differences in SCFA concentration may have been related to differences in growth recorded in the current study, especially on d 14. SCFAs can influence growth performance in different ways. Firstly, as previously mentioned, SCFA can be used as an energy source for colonic cells, which increases the nutrients available to the host for growth. Secondly, xylanase enzymes randomly cleave NSP, reducing digesta viscosity and increasing nutrient absorption resulting in a decrease in the amount of nutrients available to the micro-organisms the caeca for fermentation (Lee *et al*., 2017).

Accepted for publication 22 August 2019

One of the most notable detrimental nutrients fermented by colonic bacteria is protein. Protein fermentation by colonic bacteria has been associated with the production of toxic compounds, such as ammonia, which can inhibit growth and even cause disease (Apajalahil and Vinenola, 2016). To combat this, it has been recommended that xylanases should be used to increase nutrient utilisation and to create fermentable carbohydrates (Apajalahil and Vinenola, 2016). The current data agreed with this, as growth performance and SCFA production increased following the xylanase supplementation, however there was no effect on nitrogen digestibility.

Pre- caecal NSP fraction concentration

NSP concentration is a way to measure its hydrolysis by giving an indication of the resulting sugars in the liquid and solid phase of the ileal digesta. In a previous study, WU NSP concentration decreased while WE NSP concentration increased following xylanase supplementation (Olukosi *et al.,* 2015). A reduction in NSP concentration indicates greater hydrolysis, which is beneficial when xylanase is used to reduce digesta viscosity. In the current study, however, the aim was to increase the concentration of potentially prebiotic oligosaccharides and investigate their effect on growth performance. NSP from wheat contains large amount of arabinoxylan (AX).

WE NSPs are the main cause of increased digesta viscosity in broilers, which is counteracted by using carbohydrases such as xylanase (Choct, 2015). The concentration of WE NSP in the ileum of broilers during the current study increased when either xylanase or XOS were supplemented in the diets. This is in agreement with previous published data, as it has been reported that the disappearance of NSP from the digestive tract of broilers was significantly affected by enzyme addition (Cowieson *et al*., 2016; Cozannet *et al.,* 2017). From the data above, fructose and

Accepted for publication 22 August 2019

galactose concentration (which likely represented fructo- oligosaccharide (FOS and galacto- oligosaccharide (GOS) or galactans) was significantly increased due to xylanase addition. In addition to this, xylose concentration tended to increase XOS addition.

This tendency for xylose to increase following XOS supplementation could be as a result of undigested XOS bypassing digestion, but stoichiometry suggested this could not have been the only source of xylan in the WE fraction as 0.1 g/kg was added to the diet, and concentration increased by 3 g/ kg DM. An increase in galactose (representing galactans and pectins) concentration in the ileum of broilers following carbohydrase supplementation has been reported previously (Kocher *et al.,* 2002). This is of interest, because FOS and GOS have been associated with prebiotic effects (Kaplan and Hutkins, 2000; Boehm *et al.,* 2005; Courtin *et al.,* 2008).

WU NSP are generally not fermented, unlike WE NSP; however, they have been associated with gut development (Choct, 2015). In the current study, the concentration of arabinose, fructose and galactose were increased in response to xylanase or XOS supplementation in the WU NSP fraction. This could suggest that xylanase may have beneficial effects on gut development which is in agreement with other studies (Jimenez –Monero *et al.,* 2009; Liu *et al.,* 2017). This is certainly true for the gizzard, where increases in dietary fibre have been associated with heavier gizzards, resulting in increased retention time of digesta and smaller particle size which, in turn, increases nutrient digestibility (Jimenez- Monero *et al.,* 2009). The increase in WU NSP concentration could improve the development of the gastrointestinal tract, contributing to improvements in growth performance.

Accepted for publication 22 August 2019

The current trial data demonstrated similarities between the effects of xylanase and XOS supplementation on growth performance, pre-caecal NSP concentration and caecal SCFA concentration, indicating similar modes of action. Consequently, it can be suggested that improvements in growth performance were partly driven by the production of in-situ prebiotics following xylanase supplementation. This study suggested that supplementing broiler diets with low levels of xylanase or high levels of XOS had a positive effect on the concentration of WU NSP fractions in the ileum, however, there was a limited effect of high levels of xylanase or XOS on performance. As such, more research into increasing the inclusion level of xylanase or XOS is required.

Acknowledgements

The authors acknowledge the staff and students at the Monogastric Science Research Centre, Ayr for all of their help and support.

Allison Craig is a recipient of i-CASE PhD studentship from Biotechnology and Biological Sciences Research Council (BBSRC). SRUC receives funding from the Scottish government (RESAS).

Disclosure statement

M.R Bedford is an employee of AB Vista, the manufacturer of the enzymes used in this study.

References

Accepted for publication 22 August 2019

ABDEL- HAFEEZ, H.M, E.S.E. SALEH, S. S.TAWFEEK, I.M.I YOUSSEF and A.S.A ABDEL- DAIM. 2017. "Effects of Probiotic, Prebiotic and Symbiotic With and Without Feed Restriction on Performance, Haematological Indices and Carcass Characteristics of Broiler Chickens". *Asian- Australasian Journal of Animal Science* 30: 672-682.

AL-SULTAN, S., S.M. ABDEL- RAHEEM , W.R. EL- GHAREEB and M.H.A MOHAMED. 2016."Comparative Effects of Using Prebiotic, Probiotic, Symbiotic and Acidifier on Growth Performance, Intestinal Microbiology and Histomorphology of Broiler Chicks". *Japanese Journal of Veterinary Research* 64 (supplement 2): S187-195.

APAJALAHIL, J., and K. VINENOLA. 2016. "Interaction Between Chicken Intestinal Microflora and Protein Digestion". *Animal Feed Science and Technology* 221: 323-330**.**

ARSI, K, A.M DONOGUE, A. WOO-MING, P.J. BLORE and D.J DONOGUE. 2015. "The Efficacy of Selected Probiotic and Prebiotic Combinations in Reducing Campylobacter Colonisation in Broiler Chickens". *Journal of Applied Poultry Research* 24: 327-334.

AOAC 2006. "Official Methods for Analysis, 18th ed". *Association of Official Analytical Chemists, Washington, DC.*

BOEHM, G, B. STAHL, J. JELINEK, J. KNOL, V. MINIELLO and G.E. MORO. 2005.

Accepted for publication 22 August 2019

"Prebiotic Carbohydrates in Human Milk and Formulas". *Aceta Paediatrica* 94 (suppl 449): 18-21.

CHOCT, M. 2015. "Feed Non-Starch Polysaccharides for Monogastric Animals: Classification and Function". *Animal Production Science*. 27: 855-861.

COURTIN, C.M, W.F. BROEKAERT, K. SWENEN,O. LESCROART, O. ONAGBESAN, J. BUYSE, E. DECUYPERE, T. VAN DEWIELE, M. MARZORATI, W. VERSTRATE, G. HUYGHEBAERT and J.A. DELCOUR. 2008. "Dietary Inclusion of Wheat Bran Ararbinoxylooligosacchairdes Induces Beneficial Nutritional Effects in Chickens". *Cereal Chemistry*. 85: 607-613.

COWIESON, A.J, H. LU, K.M. AJUWON, I. KNAP and O. ADEOLA. 2017. "Interactive Effects of Dietary Protein Source and Exogenous Protease and Jejunal Health of Broiler Chickens". *Animal Production Science.* 57: 252-261.

COWIESON, A.J, W. SCHLIFKA, I. KNAPP, F.F. ROOS, R. SCHOOP and J.W. WILSON. 2016. "Meta- Analysis off Effect of a Mono-Component Xylanase on the Nutritional Value of Wheat Supplemented with Exogenous Phytase for Broiler Chickens". *Animal Production Science*. 56: 2014- 2022.

COZANNET, P, M.T KIDD, R.M. NETO and P.A. GERAERT. 2017. "Next-Generation Non-Starch- Polysaccharide- Degrading, Multi Carbohydrase Complex Rich in Xylanase and Arabinfuranosidase to Enhance Feed Digestibility". Poultry Science. 96: 2743-2750.

De MEASSCHALCK, C, V. EECKHAUT, L. MAERTENS, L. DE LAUGE, L. MARCHAL, C. NEZER, S. De BAERE, S. CROUBLES, G. DAUBE, J. DEWULF, F. HAESEBROUCK, R. DUCATELLE, B. TAMINAU and F.

Accepted for publication 22 August 2019

VAN IMMRSEEL. 2015. “Effects of Xylo-oligosaccharides on Broiler Chicken Performance and Microbiota”. *Applied and Environmental Microbiology* 81: 5880-5888.

ENGBERG, R.M, M.S. HEDENANN, S.STEENFELDT AND B.B JENSEN. 2004. “Influence of Whole wheat and Xylanase on Broiler Performance and Microbial Composition and Activity in the Digestive Tract”. *Poultry Science* 82: 925-938.

ENGLYST, H. N., M.E. QUIGLEY AND G. J. HUDSON. 1994. “Determination of Dietary Fibre as Non-Starch Polysaccharides with Gas-Liquid Chromatographic, High-Performance Liquid Chromatographic or Spectrophotometric Measurement of Constituent Sugars”. Analysts 119: 1497–1509.

FRANCESCH. M AND P.A GERAERT. 2009. “ Enzyme Complex Containing NSPase and Phytase Improves Growth Performance and Bone Mineralisation of Broilers Fed Reduced Nutrient Cron-Soybean- Based Diets”. *Poultry Science* 88; 1915-1924.

GAO,F., Y. JIANG, G.H. ZHOU, AND Z.K. HAN. 2007. “The Effects of Xylanase Supplementation on Growth, Digestion, Circulating Hormone and Metabolite Levels, Immunity and Gut Microflora in Cockerels Fed on Wheat Based Diets”. *British Poultry Science* 48: 480-488.

GONG J, H. YU, T. LIU, J.J GILL, J.R. CHAMBERS, R. WHEATCROFT AND P.M. SABOUR. 2008. “Effects of Zinc Bacitracin, Bird Age and Access to Range on Bacterial Microbiota in the Ileum and Caeca of Broiler Chickens”. Journal of Applied Microbiology 104 :1372-1382.

Accepted for publication 22 August 2019

GONZALEZ- ORTIZ, G, O. OLUKOSI AND M.R BEDFORD. 2016. "Evaluation of the Effect of Different Wheats and Xylanase Supplementation on Performance, Nutrient and Energy Utilisation in Broiler Chicks". *Animal Nutrition* 2; 173-179.

GOVIL, K., S. NAYAK, R.P.S BAGHEL, A.K. PATIL, C.D.MALAPURE and D. THAKUR. 2017. "Performance of Broiler Chicken Fed Multi-Carbohydrases Supplemented Low Energy Diet". *Veterinary World* 10: 727-731.

JIA, W., B.A. SLOMINSKI, H.L. BRUCE, G. BLANK, G. CROW and O. JONES. 2009. "Effects of Diet Type and Enzyme Addition on Growth Performance and Gut Health of Broiler Chickens During Sub Clinical *Clostridium perfingens* Challenge". *Poultry Science* 88: 132-140.

JIMENEZ- MONERO, E, J.M. GINZALEZ-ALVARADO, A. DECOCA- SINOVA, R. LAZARO and G.G. MATEOS. 2009. "Effects of Source of Fibre on the Development and pH of the Gastrointestinal Tract of Broilers". *Animal Feed Science and Technology* 154: 93-101.

JUNG, S.J, R. HOUDE, B. BAURHOO, X. ZHAO and B.H. LEE. 2008. "Effects of Galacto- oligosaccharides and a Bifidobacteria Lactic-based Probiotic Strain on the Growth Performance and Faecal Microflora of Broiler Chickens". *Poultry Science* 87: 1694-1699.

KAMRAN, Z., M. SARWAR, M. NISA, M.A. NADEEM, S.MAHMOOD, M.E.BABAR, and S. AHMED. 2008. "Effect of Low Protein Diets Having Constant Energy-to- Protein Ratio on Performance and Carcass Characteristics of

Broiler Chickens From One to Thirty- Five Days of Age". *Poultry Science* 87: 468-474.

KAPLAN, H and R.W. HUTKINS. 2000. "Fermentation of Fructooligosaccharides by Lactic Acid Bacteria and Bifidobacteria". *Applied and Environmental Microbiology* 66: 2682-2684.

KHATTAK, F, V. PASCHALIS, M. GREEN, J.G.M HOUDJIK, P. SOULTANAS AND J. MAHDAVI. 2018. "TYPLEX® Chelate, a Novel Feed Additive, Inhibits *Camplybacter jejuni* Biofilm Formation and Cecal Colonization in Broiler Chickens". *Poultry Science* 97: 1319-1399.

KIARIE, E, L.F. ROMERO and V. RAVINDRAN. 2014. "Growth Performance, Nutrient Utilisation and Digesta Characteristics in Broiler Chickens Fed Corn or Wheat Diets Without or With Supplemental Xylanase". *Poultry Science* 93: 1186-1196.

KIRKPINAR, K, Z. ACIKGOZ, M. BOZKURT and V. AYHAN. 2004. "Effects of Inclusion of Poultry By-product Meal and Enzyme- Prebiotic Supplementation in Grower Diets on Performance and Feed Digestibility of Broilers". *British Poultry Science* 45: 273-279.

KNUDSEN, K.E.B. 2014. "Fibre and Non-Starch Polysaccharide Content and Variation in Common Crops used in Broiler Diets". *Poultry Science* 93: 2380-2393.

KOCHER, A, M. CHCOT, M.D. PORTER and J. BROZ. 2002. "Effect of Feed Enzymes on the Nutritive Value of Soybean Meal Fed to Broilers". *British Poultry Science* 43: 54-63.

Accepted for publication 22 August 2019

LEE, S.A, J. APAJALAHTI, K. VIENOLA, G. GONZALEZ-ORTIZ, C.M.G.A. FONTES and M.R. BEDFORD. 2017. "Age and Dietary Xylanase Supplementation Affects Ileal Sugar Residues and Short Chain Fatty Acid Concentration in the Ileum and Caecum of Broiler Chickens". *Animal Feed Science and Technology* 234: 29-42.

LENTLE, R.G. 2005. "The Macro-biophysics of Digestion; Implications for the Poultry Industry". *Australian Poultry Science Symposium* 17: 163-170.

LIU, W.C and I.H. KIM. 2017. "Effects of Dietary Xylanase Supplementation on Performance and Functional Digestive Parameters in Broilers fed Wheat-based Diets". *Poultry Science* 96: 566-573.

MATHLOUTHI, N, S. MALLET, L. SAULINER, B. QUEMENER and M. Larbier. 2002. "Effects of Xylanase and β-glucanase Addition on Performance, Nutrient Digestibility and Physico Chemical Conditions in the Small Intestine Contents fed a Wheat and Barley Based Diet". *Animal Research* 51: 395- 406.

MENG, X. and B.A. SLOMINSKI. 2005."Nutritive Values of Corn, Soybean Meal, Canola Meal and Peas for Broiler Chickens as Affected by a Multicarbohydrase Preparation of Cell Wall Degrading Enzymes". *Poultry Science* 84: 1242-1251.

MOUNTZOURIS, K. C., P. TSITRSIKOS, I. PALAMIDI, A. ARVANITI, M. MOHNL, G. SCHATZMAYR and K. FEGEROS. 2010. "Effects of Probiotic Inclusion Levels in Broiler Nutrition on Growth Performance, Nutrient Digestibility, Plasma Immunoglobulins, and Caecal Microflora Composition". *Poultry Science* 89**:** 58-67.

Accepted for publication 22 August 2019

OLUKOSI, O. A., A.J. COWIESON and O. ADEOLA. 2007. "Age-related Influence of a Cocktail of Xylanase, Amylase, and Protease or Phytase Individually or in Combination in Broilers". *Poultry Science* 86: 77–86.

OLUKOSI, O.A, L.A. BEESON, K. ENGLYST and L.F. ROMERO. 2015. "Effects of Proteases Without or With Carbohydrases on Nutrient Digestibility and Disappearance of Non- Starch Polysaccharides in Broiler Chickens". *Poultry Science* 94: 2662-2669.

RAVANGARD, A.H, M. HOUSHMAND, M. KHAJVI AND R. NAGHIHI. 2017. "Performance and cecal bacteria counts of broilers fed low protein diets with and without a combination of probiotic and prebiotic". *Brazilian Journal of Poultry Science*. Special Issue- Nutrition: 75-82

SHORT, F.J, P. GORTON, J. WISEMAN and K.N. BOORMAN. 1996. "Determination of Titanium Oxide Added as an Inert Marker in Chicken Digestibility Studies". *Animal Feed Science Technology* 59: 215-221.

SMIRICKY- TJARDES, M.R, C.M. GRIESSHOP, E.A. FLICKINGER, L.L BAUER and G.C. Fahey Jr. 2003. "Dietary Galactooligosaccharides Affect Ileal and Total Tract Nutrient Digestibility, Ileal and Faecal Bacterial Concentrations and Ileal Fermentative Characteristics of Growing Pigs". *Journal of Animal Science* 81: 2535-2545.

WANG, Z.R, S.Y QIAO, W.Q LU and D.F LI. 2005. "Effects of enzyme

Accepted for publication 22 August 2019

supplementation on performance, nutrient digestibility, gastrointestinal morphology and volatile fatty acid profiles in the hindgut of broilers fed wheat based diets". *Poultry Science* 84: 875-881.

WILLIAMS, M.P, J.T. KLEIN, C.L. WYATT, T.W. YORK and J.T. Lee. 2014. "Evaluation of Xylanase in Low Energy Broiler Diets". *Journal of Applied Poultry Research* 23: 188-195.

XU, Z.R., G.H. HU, M.S. XIA, X.A. ZHAN and M.Q. Wang. 2003. "Effects of Dietary Fructooligosacchairde on Digestive Enzyme Activities, Intestinal Microflora and Morphology of Male Broilers". *Poultry Science* 82: 1030-1036.

YANG, Y, P.A. IJI, L.L MIKKELSEN and M. Choct. 2008. "Effects Dietary Mannanoligosacchairde on Growth performance and Gut Development of Broilers Given Different Cereal- Based Diets". *Journal of Animal Physiology and Animal Nutrition* 92: 650-659.

YOUSAF, M.S, A. IJAZ, K. ASHRQAF, M.A. RASHID, A. HAFEEZ, H. ZANEB, E. DAR, R. NASEER, I. RABBANI, J. ZENTEK and H. Reham. 2016. "Comparative Effects of Different Dietary Concentrations of β-galacto-oligosaccharides on Growth Performance, Feed Efficiency and Organ Development in Broiler Chickens". *The Journal of Animal and Plant Science* 26: 1603-1608.

ZHANG, L, J. XU, L. LEI, Y. JIANG, F. GAO and G.H ZhOU. 2014. "Effects of Xylanase Supplementation on Growth Performance, Nutrient Digestibility and Non-Starch Polysaccharide Degradation in Difference Sections of the

Accepted for publication 22 August 2019

Gastro-intestinal Tract of Broilers". *Asian-Australasian Journal of Animal Science* 27: 855-861.

ZHENPING, S, L WENTING, Y RUIKUI, L JIA, L HONGHONG, S WEI, W ZHONGMIE, L JINGPAN, S ZHE and Q YULING. 2013. "Effect of straw derived oligosaccharide on broiler growth performance, endocrine metabolism and immune response". *Canadian Journal of Veterinary Research* 77: 105-109.

Accepted for publication 22 August 2019

Table 1. The calculated and analysed nutrient content of wheat-based diets supplemented with enzymes or prebiotic oligosaccharide.

Ingredient (g/kg)	Control
Wheat	556.5
Wheat Bran	192.5
Soybean meal	152.5
Soya oil	25.0
Limestone	12.5
Dicalcium Phos, 18%P	14.0
Sodium Bicarbonate	6.0
Lysine HCl	5.0
Methionine	2.0
Threonine	2.0
Valine	2.0
Vitamin & Mineral premix	5.0
TiO2 Marker	25.0
Total	1000.0
Calculated content	
ME (MJ/kg)	11.41
Crude Protein (g/kg)	200.0
Calcium (g/kg)	10.0
Phosphorus (g/kg)	7.5
nPP (g/kg)	4.9
Na (g/kg)	2.1
Cl (g/kg)	3.5
Analysed nutrient content	
DM (g/kg)	877.1
AME (MJ/kg)	16.19
Na (g/kg)	1.9
Cl (g/kg)	3.1
Calcium (g/kg)	9.6
Phosphorus (g/kg)	6.3
N(g/kg)	31.3

Notes; TiO2- titanium dioxide; Na- sodium; Cl- chloride; DM- dry matter; AME- apparent metabolizable energy; N- nitrogen

Accepted for publication 22 August 2019

Table 2. The growth performance of broilers fed diets deficient in energy and protein and supplemented with xylanase or XOS.

		Day 14			Day 28		
Additive	Inclusion Level	BWG (g/bird)	FI (g/bird)	FCR	BWG (g/bird)	FI (g/Bird)	FCR
No Additive		388.1	596.2	1.54	1313.3	2953.9	2.26
Xylanase	Low	381.1	594.1	1.56	1294.5	2707.1	2.10
	High	394.3	579.2	1.47	1320.0	2546.5	1.93
XOS	Low	356.7	561.5	1.59	1261.6	2536.5	2.03
	High	375.1	561.7	1.51	1306.4	2604.7	2.00
	SEM	10.449	12.08	0.038	29.202	86.161	0.078
P-value of control vs additive[1]		0.390	0.111	0.752	0.592	<0.001	0.009
Means for main effect of additive type (AT)							
Xylanase		387.7[b]	586.7[b]	1.52[a]	1307.3	2626.8	2.02
XOS		365.9[a]	561.6[a]	1.55[b]	1284.0	2570.6	2.02
	SEM	10.449	12.08	0.038	29.202	86.161	0.078
Means for main effect of inclusion level (IL)							
	Low	368.9	577.8	1.57	1278.0	2621.8	2.07
	High	384.7	570.4	1.49	1313.2	2570.6	1.97
	SEM	10.449	12.08	0.038	29.202	86.161	0.078
P values							
Additive Type (AT)		0.044	0.045	0.449	0.430	0.518	0.978
Inclusion Level (IL)		0.139	0.543	0.034	0.236	0.595	0.197
AT × IL		0.806	0.535	0.944	0.744	0.193	0.385

Notes; XOS- xylo-oligosaccharide; BWG- body weight gain; FI- feed intake; FCR; feed conversion ratio; [1]P- values for control vs. additive types - the mean for the control group was compared to all other treatments containing an additive irrespective of type or level ; [ab] different superscripts with the same column indicate means that are significantly (P<0.05) different

Accepted for publication 22 August 2019

Table 3. Coefficients of energy and nitrogen digestibility of broilers aged 29 days and fed diets supplemented with xylanase or XOS

Additive	Inclusion Level	DM	IDE (MJ/kg)	N	Energy Intake (units/bird)	N Intake (units/bird)
No Additive		0.713	13.77	0.858	38.44	85.37
Xylanase	Low	0.652	13.28	0.837	33.23	81.18[b]
	High	0.606	11.99	0.817	29.62	69.15[a]
XOS	Low	0.630	12.22	0.812	29.54	70.23[ab]
	High	0.624	12.23	0.821	30.26	79.64[ab]
	SEM	0.0189	0.372	0.00913	1.499	1.987
P-value of control vs additive[1]		<0.001	0.003	0.001	<0.001	0.002
Means for main effect of additive type (AT)						
Xylanase		0.629	12.63	0.827	31.39	75.71
XOS		0.627	12.23	0.816	29.94	74.39
	SEM	0.0189	0.263	0.00646	1.06	1.987
Means for main effect of inclusion level (IL)						
	Low	0.641	12.75	0.824	31.43	75.16
	High	0.615	12.11	0.819	29.90	74.94
	SEM	0.0189	0.263	0.00646	1.06	1.405
P values						
Additive Type (AT)		0.170	0.095	0.550	0.317	0.937
Inclusion Level (IL)		0.909	0.284	0.245	0.343	0.643
AT × IL		0.302	0.090	0.115	0.158	<0.001

Notes; XOS- xylo-oligosaccharide; DM- dry matter; IDE- ileal digestible energy; N- nitrogen; [1]P- values for control vs. additive types - the mean for the control group was compared to all other treatments containing an additive irrespective of type or level; [abc] different superscripts with the same column indicate means that are significantly (P<0.05) different

Accepted for publication 22 August 2019

Table 4. The effect feeding wheat-based diets supplemented with xylanase or XOS on SCFA concentrations (mg/kg) in the caeca on days 14 and 28

Additive	Inclusion Level	Day 14 Acetic Acid	Propionic Acid	n- butyric Acid	n- Valeric Acid	Total SCFA	Day 28 Acetic Acid	Propionic Acid	n- butyric Acid	n- Valeric Acid	Total SCFA
No Additive		4749	75.70	1260.7	45.3	6242	4790	193	1046	69.1	6170
Xylanase	Low	4260	56.60	1117.8	23.1	5532	5284	198	1347	78.2	6984
	High	4036	61.0	1250.0	28.7	6438	5541	172	1563	80.8	7435
XOS	Low	4949	112.0	1348.6	40.6	6528	5374	166	1345	80.1	7056
	High	5244	136.4	1505.9	55.0	7018	5177	222	1194	82.9	6756
Pooled SEM		108.8	35.54	143.13	9.87	437.2	234.9	28.9	114.8	6.92	310.6
P-value of control vs. Additive[1]		0.737	0.066	0.874	0.596	0.818	0.042	0.922	0.019	0.149	0.015
Means for main effect of additive types (AT)											
Xylanase		4148[a]	56.80[a]	1183.9	32.4[a]	5485[a]	5412	185	1455	79.5	7210
XOS		5096[b]	124.2[b]	1427.3	47.8[b]	6773[b]	5275	194	1269	81.5	6906
Pooled SEM		236.5	47.66	172.07	10.89	309.1	116.1	20.5	81.1	4.89	219.6
Means for main effect of inclusion level (IL)											
	Low	4605	82.30	1233.2	38.4	6030	5412	185	1455	79.5	7210
	High	4640	98.70	1378.0	41.9	6228	5275	194	1269	81.5	6906
Pooled SEM		236.5	11.60	102.35	2.47	309.1	116.1	20.5	81.1	4.89	219.6
P- values for main effects and interactions											
Additive Type (AT)		0.007	<0.001	0.128	0.026	0.006	0.563	0.760	0.114	0.774	0.335
Inclusion Level (IL)		0.917	0.265	0.335	0.759	0.654	0.899	0.612	0.778	0.699	0.808
AT × IL		0.442	0.546	0.789	0.234	0.509	0.341	0.168	0.118	0.989	0.234

Accepted for publication 22 August 2019

Notes; XOS- xylo-oligosaccharide; [1]P- values for control vs. additive types - the mean for the control group was compared to all other treatments containing an additive irrespective of type or level; [ab] different superscripts with the same column indicate means that are significantly (P<0.05) different

Table 5. The effect of feeding wheat-based diets supplemented with xylanase or XOS on the concentration (g/100g) of WU NSP fractions in the ileum of broilers

Additive	Inclusion Level	Rhamnose	Fructose	Arabinose	Xylose	Galactose	TOTAL (g/100g)
No Additive		0.0156	0.0184	1.647[a]	2.54	0.583[a]	8.74
Xylanase	Low	0.0129	0.0170	1.721[a]	2.65	0.572[a]	9.01
	High	0.0222	0.0374	2.431[b]	3.55	0.851[b]	12.13
XOS	Low	0.0261	0.0256	2.002[ab]	3.08	0.686[ab]	10.30
	High	0.0434	0.0450	1.950[ab]	2.90	0.688[ab]	10.35
	Pooled SEM	0.004	0.005	0.155	0.292	0.047	0.840
P-value of control vs additive types[1]		0.022	0.044	0.042	0.142	0.039	0.086
Means for main effect of additive type (AT)							
Xylanase		0.0176[a]	0.0272	2.076	3.10	0.711	10.57
XOS		0.0347[b]	0.0353	1.976	2.99	0.687	10.32
	Pooled SEM	0.003	0.004	0.110	0.206	0.033	0.594
Means for main effect of inclusion level							
	Low	0.0195[a]	0.0213[a]	1.861[a]	2.87	0.629[a]	9.65
	High	0.0328[b]	0.0412[b]	2.190[b]	3.22	0.770[b]	11.24
	Pooled SEM	0.003	0.004	0.110	0.206	0.033	0.594
P- values for main effects and interactions (IL)							
Additive Type (AT)		<0.001	0.144	0.525	0.717	0.608	0.776
Inclusion Level (IL)		0.002	0.001	0.047	0.235	0.007	0.074
AT × IL		0.302	0.937	0.024	0.083	0.008	0.084
P- Values for Contrasts							
Control vs. Xylanase, low level				0.739		0.864	
Control vs. Xylanase, high level				0.002		<0.001	
Control vs. XOS, low level				0.122		0.139	

Accepted for publication 22 August 2019

Control vs. XOS high level	0.184	0.130

Notes; XOS- xylo-oligosaccharide; GlucA2- glucuronic acid; [1]P-value for control vs. additive types- the mean for the control group was compared to all other treatments containing an additive irrespective of type or level; [ab] different superscripts within the same column indicate means that are significantly different

Accepted for publication 22 August 2019

Table 6. The effect of feeding wheat based diets supplemented with xylanase or XOS on the concentration (g/100g) of WE NSP fractions in the ileum of broilers

Additive	Inclusion Level	Rhamnose	Fructose	Arabinose	Xylose	Galactose	TOTAL (g/100g)
No Additive		0.0390	0.0462	0.605	0.948	0.398	2.74
Xylanase	Low	0.0422	0.0479[a]	0.624	0.898	0.463	3.04
	High	0.0619	0.0692[b]	0.855	1.143	0.607	3.84
XOS	Low	0.0484	0.0682[b]	0.773	1.284	0.507	3.74
	High	0.0520	0.0570[a]	0.810	1.216	0.556	3.63
Pooled SEM		0.010	0.007	0.08	0.115	0.052	0.351
P-value of control vs additive types [1]		0.278	0.083	0.081	0.162	0.030	0.051
Means for main effect of additive types (AT)							
Xylanase		0.0520	0.0586	0.739	1.02	0.535	3.44
XOS		0.0502	0.0626	0.791	1.25	0.532	3.68
Pooled SEM		0.007	0.005	0.055	0.081	0.037	0.248
Means for main effect of inclusion level (IL)							
	Low	0.0453	0.0580	0.699	1.091	0.485	3.39
	High	0.0569	0.0631	0.832	1.179	0.581	3.73
Pooled SEM		0.007	0.005	0.055	0.081	0.037	0.248
P- values for main effects and interactions							
Additive Type (AT)		0.850	0.573	0.511	0.061	0.949	0.495
Inclusion Level (IL)		0.245	0.479	0.103	0.453	0.078	0.337
AT ×IL		0.471	0.032	0.227	0.190	0.369	0.261
P- values for contrasts							
Control vs. Xylanase, low level			0.866				
Control vs. Xylanase, high level			0.032				
Control vs. XOS, low level			0.039				
Control vs. XOS high level			0.291				

Accepted for publication 22 August 2019

Notes; XOS- xylo-oligosaccharide; GlucA2- glucuronic acid; [1]P-value for control vs. additive types- the mean for the control group was compared to all other treatments containing an additive irrespective of type or level; [ab] different superscripts within the same column indicate means that are significantly ($P<0.05$) different

Accepted for publication 22 August 2019